# Collective Center of ICG SB RAS "Collection of Pluripotent Human and Mammalian Cell cultures for Biological and Biomedical Research"

The collection contains embryonic stem and induced pluripotent stem cells of humans, mice and other laboratory animals. There are also immortalized cell lines and protist cultures.

Mouse embryonic stem cells can be used to produce transgenic animals.

Unique cell lines of American mink pluripotent stem cells allow studying the early embryonic development of mustelids.

Research areas:

- derivation of cell lines for fundamental and applied research in the fields of developmental biology, cell biology and transgenesis, including embryonic stem and induced pluripotent stem cells of humans, mice and other laboratory animals, primary and with genetic modifications;
- systematic description of the generated cell lines;
- quality control of collection material using modern methods.

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# Contents

# Mouse pluripotent stem cells

## Cell line passport of DGES1

**Catalogue number:** MMES00001
**Name:** DGES1
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from 129S2/SvPasCrl 3.5D blastocyst
**Authors:** Menzorov A.G., Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-41, tetraploid cells 6%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by embrioid body formation, teratoma formation in SCID mice and germ line transmission
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 6
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1002/jcb.28981

# Cell line passport of DGES2

**Catalogue number:** MMES00002
**Name:** DGES2
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from 129S2/SvPasCrl 3.5D blastocyst
**Authors:** Menzorov A.G., Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-41, tetraploid cells 5%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by embrioid body formation, teratoma formation in SCID mice and germ line transmission
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 6
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1002/jcb.28981

# Cell line passport of DGES1-TubbEGFPpuro

**Catalogue number:** MMES00038
**Name:** DGES1-TubbEGFPpuro
**Description:** DGES1 mouse embryonic stem cells with site-specific EGFP insertion after bTubb3 last exon via 2A peptide and puromycin resistance
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-41, tetraploid cells less than 10%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 24
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:**
**Date:** 01.01.2017

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1002/jcb.28981

# Cell line passport of DGES1-TubbEGFP

**Catalogue number:** MMES00039
**Name:** DGES1-TubbEGFP
**Description:** DGES1 mouse embryonic stem cells with site-specific EGFP insertion after bTubb3 last exon via 2A peptide
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-41, tetraploid cells less than 10%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 22
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:**
**Date:** 01.01.2017

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1002/jcb.28981

# Cell line passport of DGES1-TubbEGFPSV40puro

**Catalogue number:** MMES00040
**Name:** DGES1-TubbEGFPSV40puro
**Description:** DGES1 mouse embryonic stem cells with site-specific EGFP insertion after bTubb3 last exon via 2A peptide, SV40 polyA signal and puromycin resistance (site-specific and unspecific insert)
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-41, tetraploid cells less than 5%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 20
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:**
**Date:** 01.01.2017

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1002/jcb.28981

# Cell line passport of DGES1-TubbEGFPSV40

**Catalogue number:** MMES00041
**Name:** DGES1-TubbEGFPSV40
**Description:** DGES1 mouse embryonic stem cells with site-specific EGFP insertion after bTubb3 last exon via 2A peptide, SV40 polyA signal and puromycin resistance (unspecific insert)
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-40, tetraploid cells less than 2%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 26
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:**
**Date:** 01.01.2017

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1002/jcb.28981

# Cell line passport of MA01

**Catalogue number:** MMES00004
**Name:** MA01
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-45, tetraploid cells less than 2%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by teratoma formation in NU/NU mice and germ line transmission
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA01-3E

**Catalogue number:** MMES00037
**Name:** MA01-3E
**Description:** MA01 mouse embryonic stem cells with EGFP cassette in Trim71 gene
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-41, tetraploid cells less than 2%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by teratoma formation in SCID mice and germ line transmission
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 15
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:**
**Date:** 01.01.2017

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA02

**Catalogue number:** MMES00005
**Name:** MA02
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 39-41, tetraploid cells 13%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by teratoma formation in NU/NU mice and chimeric animal generation
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA03

**Catalogue number:** MMES00006
**Name:** MA03
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 40-42, 71-83, tetraploid cells 79%, modal chromosome number 78, 79
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 7
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA04

**Catalogue number:** MMES00007
**Name:** MA04
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 39-42, tetraploid cells 28%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by teratoma formation in NU/NU mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA05

**Catalogue number:** MMES00008
**Name:** MA05
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-42, tetraploid cells 18%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 8
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA06

**Catalogue number:** MMES00009
**Name:** MA06
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 40-41, tetraploid cells 17%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by teratoma formation in NU/NU mice and chimeric animal generation
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA07

**Catalogue number:** MMES00010
**Name:** MA07
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 39-43, tetraploid cells 24%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA08

**Catalogue number:** MMES00011
**Name:** MA08
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 39-41, tetraploid cells 11%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA09

**Catalogue number:** MMES00012
**Name:** MA09
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-41, tetraploid cells 8%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA10

**Catalogue number:** MMES00013
**Name:** MA10
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 39-43, tetraploid cells 7%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA11

**Catalogue number:** MMES00014
**Name:** MA11
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 39-41, tetraploid cells 20%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 7
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA12

**Catalogue number:** MMES00015
**Name:** MA12
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-42, tetraploid cells 2%, modal chromosome number 40, 41
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 5
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA13

**Catalogue number:** MMES00016
**Name:** MA13
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, X0, chromosome number variability 39-43, tetraploid cells 8,6%, modal chromosome number 39
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MA15

**Catalogue number:** MMES00017
**Name:** MA15
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from outbred BALB x 129/Ola blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-45, tetraploid cells 5%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by teratoma formation in NU/NU mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC01

**Catalogue number:** MMES00048
**Name:** MC01
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 39-45, tetraploid cells 32%, modal chromosome number 41, 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC02

**Catalogue number:** MMES00049
**Name:** MC02
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-43, tetraploid cells 16%, modal chromosome number 41, 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 5
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC03

**Catalogue number:** MMES00050
**Name:** MC03
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-45, tetraploid cells 6%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by teratoma formation in NU/NU mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 6
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC04

**Catalogue number:** MMES00051
**Name:** MC04
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 41-45, tetraploid cells 15%, modal chromosome number 42
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 5
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC05

**Catalogue number:** MMES00052
**Name:** MC05
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-40, tetraploid cells 15%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC06

**Catalogue number:** MMES00053
**Name:** MC06
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 39-41, tetraploid cells 52%, modal chromosome number 40, 78, 79
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC07

**Catalogue number:** MMES00054
**Name:** MC07
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 40-43, tetraploid cells 18%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC08

**Catalogue number:** MMES00055
**Name:** MC08
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-41, tetraploid cells 21%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC09

**Catalogue number:** MMES00056
**Name:** MC09
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 40-43, tetraploid cells 18%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC10

**Catalogue number:** MMES00057
**Name:** MC10
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 40-43, tetraploid cells 18%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 11
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC11

**Catalogue number:** MMES00058
**Name:** MC11
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 40-43, tetraploid cells 25%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by teratoma formation in NU/NU mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 6
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC12

**Catalogue number:** MMES00059
**Name:** MC12
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 39-42, tetraploid cells 8%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by teratoma formation in NU/NU mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC13

**Catalogue number:** MMES00060
**Name:** MC13
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 39-43, tetraploid cells 8%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 8
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MC15

**Catalogue number:** MMES00061
**Name:** MC15
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from C57BL x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XY, chromosome number variability 40-44, tetraploid cells 20%, modal chromosome number 41, 42, 43
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MD01

**Catalogue number:** MMES00062
**Name:** MD01
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from DD/c x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XX, chromosome number variability 39-44, tetraploid cells 26%, modal chromosome number 40
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 7
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Cell line passport of MD02

**Catalogue number:** MMES00063
**Name:** MD02
**Description:** mouse (*Mus musculus*) embryonic stem cells derived from DD/c x (outbred BALB x 129/Ola) blastocyst
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=40, XXX0, chromosome number variability 40-48, 54-60, tetraploid cells 95%, modal chromosome number 80
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 6
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:** blastocyst
**Date:** 01.01.2016

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1007/s10616-014-9751-y

# Hybrid cells

## Cell line passport of tme13

**Catalogue number:** MMHC00042
**Name:** tme13
**Description:** hybrid cells produced by mouse tau-GFP ES cell and m5S embryonic fibroblast fusion, embryonic stem cell phenotype
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 4n=80, XX0, 83% of the cells have number of chromosomes 66-79
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:**
**Date:** 01.01.2017

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1038/s41598-017-18352-4

# Cell line passport of tme14

**Catalogue number:** MMHC00043
**Name:** tme14
**Description:** hybrid cells produced by mouse tau-GFP ES cell and m5S embryonic fibroblast fusion, embryonic stem cell phenotype
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 4n=80, XX0, 92% of the cells have number of chromosomes 69-77
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 17
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:**
**Date:** 01.01.2017

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1038/s41598-017-18352-4

# Cell line passport of tme17

**Catalogue number:** MMHC00044
**Name:** tme17
**Description:** hybrid cells produced by mouse tau-GFP ES cell and m5S embryonic fibroblast fusion, embryonic stem cell phenotype
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 4n=80, XX0, 75% of the cells have number of chromosomes 71-79
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:**
**Date:** 01.01.2017

**Cell culture**
**Morphology:** mouse pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1038/s41598-017-18352-4

# Cell line passport of tmf1

**Catalogue number:** MMHC00045
**Name:** tmf1
**Description:** hybrid cells produced by mouse tau-GFP ES cell and m5S embryonic fibroblast fusion, fibroblast phenotype
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 4n=80, XXY, 90% of the cells have number of chromosomes 69-82
**Pluripotency:**
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 7
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:**
**Date:** 01.01.2017

**Cell culture**
**Morphology:** fibroblast morphology
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:** https://doi.org/10.1038/s41598-017-18352-4

# Cell line passport of tmf2

**Catalogue number:** MMHC00046
**Name:** tmf2
**Description:** hybrid cells produced by mouse tau-GFP ES cell and m5S embryonic fibroblast fusion, fibroblast phenotype
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 4n=80, XXY, 76% of the cells have number of chromosomes 71-82
**Pluripotency:**
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 6
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:**
**Date:** 01.01.2017

**Cell culture**
**Morphology:** fibroblast morphology
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:** https://doi.org/10.1038/s41598-017-18352-4

# Cell line passport of tmf5

**Catalogue number:** MMHC00047
**Name:** tmf5
**Description:** hybrid cells produced by mouse tau-GFP ES cell and m5S embryonic fibroblast fusion, fibroblast phenotype
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 4n=80, XXY, 86% of the cells have number of chromosomes 73-81
**Pluripotency:**
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 6
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Mus musculus*
**Tissue:**
**Date:** 01.01.2017

**Cell culture**
**Morphology:** fibroblast morphology
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:** https://doi.org/10.1038/s41598-017-18352-4

# American mink pluripotent stem cells

## Cell line passport of MES12

**Catalogue number:** NVES00003
**Name:** MES12
**Description:** american mink (Neovison vison) embryonic stem cells derived from morula
**Authors:** Sukoyan M.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY
**Pluripotency:** pluripotency is shown by embrioid body formation and teratoma formation in immunodeficient mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 11
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** morula
**Date:** 01.01.1993

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of MES20

**Catalogue number:** NVES00018
**Name:** MES20
**Description:** american mink (Neovison vison) embryonic stem cells derived from morula
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-30, ~60, tetraploid cells 71%, modal chromosome number ~60
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** morula
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM (glucose 4.5 g/l), ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, LIF 1000 U/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.05%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of MES22

**Catalogue number:** NVES00019
**Name:** MES22
**Description:** american mink (Neovison vison) embryonic stem cells derived from morula
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-32, tetraploid cells 18%, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** morula
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of MES24

**Catalogue number:** NVES00020
**Name:** MES24
**Description:** american mink (Neovison vison) embryonic stem cells derived from morula
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-32, tetraploid cells 12%, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** morula
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of MES25

**Catalogue number:** NVES00021
**Name:** MES25
**Description:** american mink (Neovison vison) embryonic stem cells derived from morula
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XX, chromosome number variability 29-30, tetraploid cells 6%, modal chromosome number 30
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** morula
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of MES27

**Catalogue number:** NVES00022
**Name:** MES27
**Description:** american mink (Neovison vison) embryonic stem cells derived from morula
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-32, tetraploid cells 7%, modal chromosome number 30
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** morula
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of MES29

**Catalogue number:** NVES00023
**Name:** MES29
**Description:** american mink (Neovison vison) embryonic stem cells derived from morula
**Authors:** Matveeva N.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-32, tetraploid cells 6%, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** morula
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of iNV1XX1

**Catalogue number:** NVPS00066
**Name:** iNV1XX1
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XX, chromosome number variability 29-35, tetraploid cells 2%, modal chromosome number 30
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.11.2017

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.18699/LettersVJ-2022-8-10

# Cell line passport of iNV1XX2

**Catalogue number:** NVPS00067
**Name:** iNV1XX2
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XX, chromosome number variability 29-35, tetraploid cells 4%, modal chromosome number 30
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.11.2017

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.18699/LettersVJ-2022-8-10

# Cell line passport of iNV3

**Catalogue number:** NVPS00024
**Name:** iNV3
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G., Matveeva N.M., Khabarova A.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-32, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of iNV5

**Catalogue number:** NVPS00025
**Name:** iNV5
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G., Matveeva N.M., Khabarova A.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-30, tetraploid cells 7%, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of iNV6

**Catalogue number:** NVPS00026
**Name:** iNV6
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G., Matveeva N.M., Khabarova A.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-32, tetraploid cells 7%, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 6
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of iNV7

**Catalogue number:** NVPS00027
**Name:** iNV7
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G., Matveeva N.M., Khabarova A.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 30-32, tetraploid cells 10%, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of iNV9

**Catalogue number:** NVPS00028
**Name:** iNV9
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G., Matveeva N.M., Khabarova A.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-30, tetraploid cells 9%, modal chromosome number 30
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 12
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of iNV11

**Catalogue number:** NVPS00029
**Name:** iNV11
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G., Matveeva N.M., Khabarova A.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-32, tetraploid cells 9%, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 11
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of iNV13

**Catalogue number:** NVPS00030
**Name:** iNV13
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G., Matveeva N.M., Khabarova A.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-30, tetraploid cells 3%, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of iNV15

**Catalogue number:** NVPS00031
**Name:** iNV15
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G., Matveeva N.M., Khabarova A.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-32, tetraploid cells 6%, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of iNV18

**Catalogue number:** NVPS00032
**Name:** iNV18
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G., Matveeva N.M., Khabarova A.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-32, tetraploid cells 8%, modal chromosome number 30
**Pluripotency:** pluripotency is shown by expression of specific markers
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 6
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of iNV19

**Catalogue number:** NVPS00033
**Name:** iNV19
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G., Matveeva N.M., Khabarova A.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-30, tetraploid cells 8%, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 4
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Cell line passport of iNV20

**Catalogue number:** NVPS00034
**Name:** iNV20
**Description:** american mink (Neovison vison) induced pluripotent stem cells derived from embryonic fibroblasts
**Authors:** Menzorov A.G., Matveeva N.M., Khabarova A.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=30, XY, chromosome number variability 29-32, tetraploid cells 21%, modal chromosome number 30
**Pluripotency:** pluripotency is shown byteratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** study of pluripotency, developmental biology

**Source**
**Species:** *Neogale vison*
**Tissue:** fibroblasts
**Date:** 01.01.2015

**Cell culture**
**Morphology:** mink pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** a-MEM, ES cell qualified FBS 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:** https://doi.org/10.1186/1471-2164-16-S13-S6

# Tumor cell lines

## Cell line passport of NG-16

**Catalogue number:** HSPS00092
**Name:** NG-16
**Description:** glioma cells from tumor biopsy
**Authors:** Shnaider T.A., Pristyazhnyuk I.E., Yakovleva S.A., Stupak E.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46,XX, chromosome number variability 42-48, polyploid cells 3.2%, modal chromosome number 46
**Pluripotency:**
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** pharmaceutical substance testing

**Source**
**Species:** *Homo sapiens*
**Tissue:** glioblastoma, grade IV
**Date:** 30.11.2023

**Cell culture**
**Morphology:** fibroblast-like cells
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, FBS 10%, NEAA 1%, Glutamine 1%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell passage with 0.25% Trypsin-EDTA at the ratio 1:3
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:**

# Cell line passport of NG-19

**Catalogue number:** HSPS00093
**Name:** NG-19
**Description:** glioma cells from tumor biopsy
**Authors:** Shnaider T.A., Pristyazhnyuk I.E., Yakovleva S.A., Stupak E.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46,XY, chromosome number variability 44-47, polyploid cells 5.7%, modal chromosome number 46
**Pluripotency:**
**Additional characteristics:** chromosomal instability, presense of chromosome 1 derivatives (chromosome 1 and 19 codeletion) chromosome 2 derivatives
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** pharmaceutical substance testing

**Source**
**Species:** *Homo sapiens*
**Tissue:** glioblastoma of the left frontal lobe, grade IV
**Date:** 30.11.2023

**Cell culture**
**Morphology:** fibroblast-like cells
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, FBS 10%, NEAA 1%, Glutamine 1%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell passage with 0.25% Trypsin-EDTA at the ratio 1:3
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:**

# Cell line passport of NG-20

**Catalogue number:** HSPS00094
**Name:** NG-20
**Description:** glioma cells from tumor biopsy
**Authors:** Shnaider T.A., Pristyazhnyuk I.E., Yakovleva S.A., Stupak E.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46,XY, chromosome number variability 42-48, polyploid cells 4.7%, modal chromosome number 46
**Pluripotency:**
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** pharmaceutical substance testing

**Source**
**Species:** *Homo sapiens*
**Tissue:** astrocytoma in the area of the right temporal and parietal lobes, grade 3
**Date:** 30.11.2023

**Cell culture**
**Morphology:** fibroblast-like cells
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, FBS 10%, NEAA 1%, Glutamine 1%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell passage with 0.25% Trypsin-EDTA at the ratio 1:3
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:**

# Cell line passport of NG-23

**Catalogue number:** HSPS00095
**Name:** NG-23
**Description:** glioma cells from tumor biopsy
**Authors:** Shnaider T.A., Pristyazhnyuk I.E., Yakovleva S.A., Stupak E.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=45,X0; 46,XY, chromosome number variability 40-47, polyploid cells 2%, modal chromosome number 45
**Pluripotency:**
**Additional characteristics:** chromosome Y loss
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** pharmaceutical substance testing

**Source**
**Species:** *Homo sapiens*
**Tissue:** glioblastoma
**Date:** 30.11.2023

**Cell culture**
**Morphology:** fibroblast-like cells
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, FBS 10%, NEAA 1%, Glutamine 1%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell passage with 0.25% Trypsin-EDTA at the ratio 1:3
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:**

# Cell line passport of NG-25

**Catalogue number:** HSPS00096
**Name:** NG-25
**Description:** glioma cells from tumor biopsy
**Authors:** Shnaider T.A., Pristyazhnyuk I.E., Yakovleva S.A., Stupak E.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46,XY, chromosome number variability 42-89, polyploid cells 9%, modal chromosome number 46
**Pluripotency:**
**Additional characteristics:** multiple chromosomal rearrangements
**Species control:** cytogenetic
**Cryoconservation passage:** 3
**Area of application:** pharmaceutical substance testing

**Source**
**Species:** *Homo sapiens*
**Tissue:** glioblastoma of the left frontal lobe, grade IV (NG-19 relapse)
**Date:** 30.11.2023

**Cell culture**
**Morphology:** neurosphere
**Cell culture method:** neurosphere
**Cell culture medium:** DMEM/F12, FBS 10%, NEAA 1%, Glutamine 1%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell passage with 0.25% Trypsin-EDTA at the ratio 1:3
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:**

# Cell line passport of CAR-YT

**Catalogue number:** HSCC00068
**Name:** CAR-YT
**Description:** NK-cell lymphoma, produced from NK-cell lymphoma YT by lentiviral transduction of a genetic construct coding chimeric antigen receptor with specificity to human PSMA protein
**Authors:** Gorchakov A.A., Kulemzin S.V., Belovezhets T.N., Chikaev A.N., Koval O.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 4n=92, XXYY, chromosome number variability 84-98, polyploid cells 12%, modal chromosome number 92
4n=92, XXYY, range of chromosomes number 84-98, poliploids 12%, modal chromosome number 92
**Pluripotency:**
**Additional characteristics:** cloning efficiency 20%; DNA region integrated into the genome, from 5'LTR to 3'LTR: strong constitutive promoter of the human *EF1a* gene, sequence encoding the signal peptide of the light chain of immunoglobulin kappa, fused with a sequence encoding a chimeric antigen receptor with specificity for the human PSMA protein. Further, the IRES element of cardiovirus A with a sequence encoding the gene for resistance to the antibiotic zeocin.
**Species control:** cytogenetic
**Cryoconservation passage:** 8
**Area of application:** immunology, oncology

**Source**
**Species:** *Homo sapiens*
**Tissue:** NK-cells
**Date:** 05.06.2018

**Cell culture**
**Morphology:** suspension cells, upon activation can form 3-8 cell colonies; cell morphology is from spheroid to moderately asymmetric
**Cell culture method:** cell suspension
**Cell culture medium:** IMDM (glucose 4.5 g/l, L-glutamine 4mM, HEPES 25 mM, Na-Pyruvate 1mM), FBS 10%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell culture resuspending every two days at the ratio 1:2
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 2 mln cells / ml
**Cell viability after cryoconservation:** 50%
**Additional information:** cell line is given to the collection for the depositing

**References:** https://doi.org/10.23868/201811039

# Cell line passport of CAR-YT-Lact

**Catalogue number:** HSCC00069
**Name:** CAR-YT-Lact
**Description:** NK-cell lymphoma, obtained from the NK-cell lymphoma YT by lentiviral integration of cassettes encoding a chimeric antigen receptor with specificity to the human protein PSMA and the RL2 peptide (lactaptin)
**Authors:** Gorchakov A.A., Kulemzin S.V., Belovezhets T.N., Chikaev A.N., Koval O.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 4n=92, XXYY, chromosome number variability 84-98, polyploid cells 12%, modal chromosome number 92
**Pluripotency:**
**Additional characteristics:** Cloning efficiency of 20%; two DNA regions integrated into the genome, from 5'LTR to 3'LTR: a) a strong constitutive promoter of the human *EF1a* gene, a sequence encoding the signal peptide of the kappa immunoglobulin light chain, fused to a sequence encoding a chimeric antigen receptor with specificity for the human PSMA protein. Then there is an IRES element of cardiovirus A with a sequence encoding the gene for resistance to the antibiotic zeocin. b) a strong constitutive promoter of the human *EF1a* gene, a sequence encoding the signal peptide of the copepod *Gaussia princeps* luciferase (GlucSP), fused to a sequence encoding the RL2 peptide (lactaptin), marked with a hexahistidine epitope. Next is the IRES element of cardiovirus A with a sequence encoding a transduction or transfection marker, the copGFP fluorescent protein of the copepod *Pontellina plumata*.
**Species control:** cytogenetic
**Cryoconservation passage:** 8
**Area of application:** immunology, oncology

**Source**
**Species:** *Homo sapiens*
**Tissue:** NK-cells
**Date:** 05.06.2018

**Cell culture**
**Morphology:** suspension cells, upon activation can form 3-8 cell colonies; cell morphology is from spheroid to moderately asymmetric
**Cell culture method:** cell suspension
**Cell culture medium:** IMDM (glucose 4.5 g/l, L-glutamine 4 mM, HEPES 25 mM, Na-Pyruvate 1 mM), FBS 10%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell culture resuspending every two days at the ratio 1:2
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 2 mln cells / ml
**Cell viability after cryoconservation:** 50%
**Additional information:** cell line is given to the collection for the depositing

**References:** https://doi.org/10.23868/201811039

# Cell line passport of CYTO-CAR-YT-Lact

**Catalogue number:** HSCC00070
**Name:** CYTO-CAR-YT-Lact
**Description:** modified NK-cell lymphoma, obtained from the NK-cell lymphoma YT by lentiviral integration of cassettes encoding a chimeric antigen receptor with specificity to the human protein PSMA and the RL2 peptide (lactaptin). In addition, genetic editing was carried out, affecting chromosome 12 and leading to an increase in cell cytotoxicity.
**Authors:** Gorchakov A.A., Kulemzin S.V., Belovezhets T.N., Chikaev A.N., Koval O.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 4n=92, XXYY, chromosome number variability 84-98, polyploid cells 12%, modal chromosome number 92
**Pluripotency:**
**Additional characteristics:** 20% cloning efficiency; CAR-YT-Lact cells were transduced with the GeCKO knockout library and cells exhibiting enhanced cytotoxicity towards PC3-PSMA targets were selected. The cells were then subcloned and individual clones were further analyzed. The genetic editing affects chromosome 12 and results in a biallelic deletion.
**Species control:** cytogenetic
**Cryoconservation passage:** 13
**Area of application:** immunology, oncology

**Source**
**Species:** *Homo sapiens*
**Tissue:** NK-cells
**Date:** 05.06.2019

**Cell culture**
**Morphology:** suspension cells, upon activation can form 3-8 cell colonies; cell morphology is from spheroid to moderately asymmetric
**Cell culture method:** cell suspension
**Cell culture medium:** IMDM (glucose 4.5 g/l, L-glutamine 4 mM, HEPES 25 mM, Na-Pyruvate 1 mM), FBS 10%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell culture resuspending every two days at the ratio 1:2
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 2 mln cells / ml
**Cell viability after cryoconservation:** 50%
**Additional information:** cell line is given to the collection for the depositing

**References:**

# Cell line passport of EBV-positive B lymphoblastoid cell line

**Catalogue number:** HSCC00081
**Name:** EBV-positive B lymphoblastoid cell line
**Description:** Epstein-Barr virus-induced human B-lymphoma. Derived from bone marrow aspirate of a patient diagnosed with multiple myeloma. The cell culture is a descendant of an Epstein-Barr virus-infected B-clone that displaced clonotypic MM cells over several in vitro culture passages.
**Authors:** Dolgova E.V., Pronkina N.V., Chernykh E.R, Bogachev S.S.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 4n=92, XXYY
**Pluripotency:**
**Additional characteristics:** karyotype: nuc ish (IGHx2)[200]/(CKS1B,CDKN2C)x2[200]/(DLEU,LAMP)x2[200]/(D17Z1,TP53)x2[200]. *IGH*/14q32 locus rearrangement, deletion/amplification of the *CKS1B*/1q21, *CDKN2C*/1p32 loci, *DLEU*/13q14.2 deletion, *LAMP*/13q34, *TP53*/17p13 not found in the plasma cells.
**Species control:** cytogenetic
**Cryoconservation passage:** 9
**Area of application:** cell biology, oncology

**Source**
**Species:** *Homo sapiens*
**Tissue:** bone marrow
**Date:** 29.09.2014

**Cell culture**
**Morphology:** cell suspension, within 2-6 hours of cultivation cells form spherical aggregates up to 80 µm in size. Single cells are also present in the suspension. The shape of individual cells varies from spherical to moderately irregular.
**Cell culture method:** cell suspension
**Cell culture medium:** α-MEM, 10% FBS, gentamycin 40 µg/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell culture resuspending every two days at the ratio 1:2
**Cryoconservation:** 50% FBS, 40% α-MEM, 10% DMSO
**Cryoconservation cell concentration:** 1 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:** cell line is given to the collection for the depositing

**References:** https://doi.org/10.1016/j.clml.2016.06.014; https://doi.org/10.1186/s12935-019-0842-x

# Immortalised cell lines

## Cell line passport of CHO-hCNTN6-HA

**Catalogue number:** CGOC00103
**Name:** CHO-hCNTN6-HA
**Description:** genetically modified CHO cell line (Chinese hamster ovary cells, *Cricetulus griseus*) with a constitutive expression of the human *CNTN6* gene and HA-tag
**Authors:** Yunusova A.M., Chvileva A.S., Shnaider T.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 22, chromosome number variability 16-20, modal chromosome number 19, polyploid cells 13%
**Pluripotency:**
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 22
**Area of application:** Notch signaling pathway study

**Source**
**Species:** *Cricetulus griseus*
**Tissue:** ovary
**Date:** 09.10.2024

**Cell culture**
**Morphology:** epithelial-like cells
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, FBS 10%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell passage with 0.25% Trypsin-EDTA at the ratio 1:3 - 1:10
**Cryoconservation:** 50% FBS, 40% DMEM/F12, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 90%
**Additional information:**

**References:**

# Cell line passport of CHO-hDLL1-HA

**Catalogue number:** CGOC00104
**Name:** CHO-hDLL1-HA
**Description:** genetically modified CHO cell line (Chinese hamster ovary cells, *Cricetulus griseus*) with a constitutive expression of the human *DLL1* gene and HA-tag
**Authors:** Yunusova A.M., Chvileva A.S., Shnaider T.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 22, chromosome number variability 19-22, modal chromosome number 20, polyploid cells 9%
**Pluripotency:**
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 20
**Area of application:** Notch signaling pathway study

**Source**
**Species:** *Cricetulus griseus*
**Tissue:** ovary
**Date:** 09.10.2024

**Cell culture**
**Morphology:** epithelial-like cells
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, FBS 10%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell passage with 0.25% Trypsin-EDTA at the ratio 1:3 - 1:10
**Cryoconservation:** 50% FBS, 40% DMEM/F12, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 90%
**Additional information:**

**References:**

# Cell line passport of CHO-hNOTCH1-FLAG

**Catalogue number:** CGOC00105
**Name:** CHO-hNOTCH1-FLAG
**Description:** genetically modified CHO cell line (Chinese hamster ovary cells, *Cricetulus griseus*) with a constitutive expression of the human *NOTCH1* gene and FLAG-tag
**Authors:** Yunusova A.M., Chvileva A.S., Shnaider T.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 22, chromosome number variability 18-21, modal chromosome number 20, polyploid cells 12%
**Pluripotency:**
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 23
**Area of application:** Notch signaling pathway study

**Source**
**Species:** *Cricetulus griseus*
**Tissue:** ovary
**Date:** 09.10.2024

**Cell culture**
**Morphology:** epithelial-like cells
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, FBS 10%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell passage with 0.25% Trypsin-EDTA at the ratio 1:3 - 1:10
**Cryoconservation:** 50% FBS, 40% DMEM/F12, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 90%
**Additional information:**

**References:**

# Cell line passport of CHO-hNOTCH2-FLAG

**Catalogue number:** CGOC00106
**Name:** CHO-hNOTCH2-FLAG
**Description:** genetically modified CHO cell line (Chinese hamster ovary cells, *Cricetulus griseus*) with a constitutive expression of the human *NOTCH2* gene and FLAG-tag
**Authors:** Yunusova A.M., Chvileva A.S., Shnaider T.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 22, chromosome number variability 20-21, modal chromosome number 21, polyploid cells 15%
**Pluripotency:**
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 23
**Area of application:** Notch signaling pathway study

**Source**
**Species:** *Cricetulus griseus*
**Tissue:** ovary
**Date:** 09.10.2024

**Cell culture**
**Morphology:** epithelial-like cells
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, FBS 10%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell passage with 0.25% Trypsin-EDTA at the ratio 1:3 - 1:10
**Cryoconservation:** 50% FBS, 40% DMEM/F12, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 90%
**Additional information:**

**References:**

# Human fibroblasts

## Cell line passport of NAF1nor

**Catalogue number:** HSAF00064
**Name:** NAF1nor
**Description:** human skin fibroblasts, donor age 32 years, XY
**Authors:** Gridina M.M.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XY
**Pluripotency:**
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 1
**Area of application:** developmental biology

**Source**
**Species:** *Homo sapiens*
**Tissue:** skin
**Date:** 01.01.2017

**Cell culture**
**Morphology:** fibroblast morphology
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, FBS 10%, NEAA 1%, Glutamine 1%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** passage with trypsin-EDTA 0.25%, split 1:3 - 1:4
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:**

# Bird fibroblasts

## Cell line passport of OFC1A

**Catalogue number:** PMOF00097
**Name:** OFC1A
**Description:** Great tit ovary cells with fibroblast morphology
**Authors:** Pristyazhnyuk I.E., Malinovskaya L.P.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2N,ZW, 6 pairs of macrochromosomes, 33 pairs of microchromosomes
**Pluripotency:**
**Additional characteristics:** presence of rearranged chromosome homologues to chromosomes 4, 5 or 6.
**Species control:** cytogenetic
**Cryoconservation passage:** 8
**Area of application:**

**Source**
**Species:** *Parus major*
**Tissue:** ovary
**Date:** 26.08.2022

**Cell culture**
**Morphology:** fibroblast-like cells
**Cell culture method:** monolayer
**Cell culture medium:** DMEM, FBS 10%, 2% chicken serum, NEAA 1%, Glutamine 1%, PenStrep 1%
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** cell passage with Trypsin-EDTA at the ratio 1:3
**Cryoconservation:** 50% FBS, 40% DMEM, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:** https://doi.org/10.3390/ani12131724

# Human induced pluripotent stem cells

## Cell line passport of iTAF2nor3

**Catalogue number:** HSPS00035
**Name:** iTAF2nor3
**Description:** human iPSCs derived from skin fibroblasts
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XY, chromosome number variability 45-47, tetraploid cells <1%, modal chromosome number 46
**Pluripotency:** pluripotency is shown by teratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 16
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 01.01.2017

**Cell culture**
**Morphology:** human pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, KSR 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 0,05
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:**

# Cell line passport of iTAF2nor4

**Catalogue number:** HSPS00036
**Name:** iTAF2nor4
**Description:** human iPSCs derived from skin fibroblasts
**Authors:** Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XY, chromosome number variability 45-47, tetraploid cells <1%, modal chromosome number 46
**Pluripotency:** pluripotency is shown by teratoma formation in SCID mice
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 16
**Area of application:** transgenesis, developmental biology

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 01.01.2017

**Cell culture**
**Morphology:** human pluripotent stem cell colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, KSR 15%, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage, split 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 0,05
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells.

**References:**

# Cell line passport of iCS-MCM1-2

**Catalogue number:** HSPS00072
**Name:** iCS-MCM1-2
**Description:** Human iPSCs derived from mononuclear blood cells of a patient with Cohen syndrome
**Authors:** Shnaider T.A., Khabarova A.A., Grigor'eva E.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XX, chromosome number variability 46-48, tetraploid cells 1%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** COH1-/-; chr8:g.100108653dup - reading frame shift; chr8:g.100494031G>T - nucleotide substitution in the untranslated sequence of the splice donor, which with a very high probability leads to its loss and disruption of the normal process of splicing and transcription of the gene (GRCh37/hg19)
**Species control:** cytogenetic
**Cryoconservation passage:** 15
**Area of application:** developmental biology, neurogenesis, Cohen syndrome

**Source**
**Species:** *Homo sapiens*
**Tissue:** mononuclear blood cells
**Date:** 23.11.2021

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:** https://doi.org/10.3390/cells12232702

# Cell line passport of iCS-MCM1-4

**Catalogue number:** HSPS00073
**Name:** iCS-MCM1-4
**Authors:** Shnaider T.A., Khabarova A.A., Grigor'eva E.V.
**Description:** Human iPSCs derived from mononuclear blood cells of a patient with Cohen syndrome
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XX, chromosome number variability 46-47, tetraploid cells 1%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** COH1-/-; chr8:g.100108653dup - reading frame shift; chr8:g.100494031G>T - nucleotide substitution in the untranslated sequence of the splice donor, which with a very high probability leads to its loss and disruption of the normal process of splicing and transcription of the gene (GRCh37/hg19)
**Species control:** cytogenetic
**Cryoconservation passage:** 15
**Area of application:** developmental biology, neurogenesis, Cohen syndrome

**Source**
**Species:** *Homo sapiens*
**Tissue:** mononuclear blood cells
**Date:** 23.11.2021

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iCS-MCM1-13

**Catalogue number:** HSPS00074
**Name:** iCS-MCM1-13
**Description:** Human iPSCs derived from mononuclear blood cells of a patient with Cohen syndrome
**Authors:** Shnaider T.A., Khabarova A.A., Grigor'eva E.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XX, chromosome number variability 46-47, tetraploid cells 1,5%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** COH1-/-; chr8:g.100108653dup - reading frame shift; chr8:g.100494031G>T - nucleotide substitution in the untranslated sequence of the splice donor, which with a very high probability leads to its loss and disruption of the normal process of splicing and transcription of the gene (GRCh37/hg19)
**Species control:** cytogenetic
**Cryoconservation passage:** 15
**Area of application:** developmental biology, neurogenesis, Cohen syndrome

**Source**
**Species:** *Homo sapiens*
**Tissue:** mononuclear blood cells
**Date:** 23.11.2021

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:** https://doi.org/10.3390/cells12232702

# Cell line passport of iCS-MCF2-5

**Catalogue number:** HSPS00075
**Name:** iCS-MCF2-5
**Description:** Human iPSCs derived from fibroblasts of a patient with Cohen syndrome
**Authors:** Khabarova A.A., Pristyazhnyuk I.E., Vladimirova E.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XX, chromosome number variability 46-47, tetraploid cells 0,5%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** COH1-/-; chr8:g.100514033T>C; chr8:g.100844663_100844664del (GRCh37/hg19)
**Species control:** cytogenetic
**Cryoconservation passage:** 15
**Area of application:** developmental biology, neurogenesis, Cohen syndrome

**Source**
**Species:** *Homo sapiens*
**Tissue:** mononuclear blood cells
**Date:** 23.11.2021

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:** https://doi.org/10.3390/cells12232702

# Cell line passport of iCS-MCF2-6

**Catalogue number:** HSPS00076
**Name:** iCS-MCF2-6
**Description:** Human iPSCs derived from fibroblasts of a patient with Cohen syndrome
**Authors:** Khabarova A.A., Pristyazhnyuk I.E., Vladimirova E.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XX, chromosome number variability 46-48, tetraploid cells 1%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** COH1-/-; chr8:g.100514033T>C; chr8:g.100844663_100844664del (GRCh37/hg19)
**Species control:** cytogenetic
**Cryoconservation passage:** 15
**Area of application:** developmental biology, neurogenesis, Cohen syndrome

**Source**
**Species:** *Homo sapiens*
**Tissue:** mononuclear blood cells
**Date:** 23.11.2021

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iCS-MCF2-24

**Catalogue number:** HSPS00077
**Name:** iCS-MCF2-24
**Description:** Human iPSCs derived from fibroblasts of a patient with Cohen syndrome
**Authors:** Khabarova A.A., Pristyazhnyuk I.E., Vladimirova E.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XX, chromosome number variability 46-47, tetraploid cells 1%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** COH1-/-; chr8:g.100514033T>C; chr8:g.100844663_100844664del (GRCh37/hg19)
**Species control:** cytogenetic
**Cryoconservation passage:** 15
**Area of application:** developmental biology, neurogenesis, Cohen syndrome

**Source**
**Species:** *Homo sapiens*
**Tissue:** mononuclear blood cells
**Date:** 23.11.2021

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:** https://doi.org/10.3390/cells12232702

# Cell line passport of iCS-MCF3-1

**Catalogue number:** HSPS00098
**Name:** iCS-MCF3-1
**Description:** Human iPSCs derived from fibroblasts of a patient with Cohen syndrome
**Authors:** Pristyazhnyuk I.E., Voinova V.Y., Safonova M.P., Lagarkova M.A., Volovikov E.A., Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX, chromosome number variability 46-47, tetraploid cells 6%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** compound heterozygous *VPS13B* gene variants (8:g.99766811A>G and 8:g.99859429G>A, HG38)
**Species control:** cytogenetic
**Cryoconservation passage:** 5
**Area of application:** modeling of Cohen syndrome, study of lipid transport disorders

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 20.06.2024

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iCS-MCF3-3

**Catalogue number:** HSPS00099
**Name:** iCS-MCF3-3
**Description:** Human iPSCs derived from fibroblasts of a patient with Cohen syndrome
**Authors:** Pristyazhnyuk I.E., Voinova V.Y., Safonova M.P., Lagarkova M.A., Volovikov E.A., Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX, chromosome number variability 46-47, tetraploid cells 2%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** compound heterozygous *VPS13B* gene variants (8:g.99766811A>G and 8:g.99859429G>A, HG38)
**Species control:** cytogenetic
**Cryoconservation passage:** 5
**Area of application:** modeling of Cohen syndrome, study of lipid transport disorders

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 20.06.2024

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iCS-MCF3-5

**Catalogue number:** HSPS00100
**Name:** iCS-MCF3-5
**Description:** Human iPSCs derived from fibroblasts of a patient with Cohen syndrome
**Authors:** Pristyazhnyuk I.E., Voinova V.Y., Safonova M.P., Lagarkova M.A., Volovikov E.A., Menzorov A.G.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX, chromosome number variability 46-48, tetraploid cells 0%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** compound heterozygous VPS13B gene variants (8:g.99766811A>G and 8:g.99859429G>A, HG38)
**Species control:** cytogenetic
**Cryoconservation passage:** 5
**Area of application:** modeling of Cohen syndrome, study of lipid transport disorders

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 20.06.2024

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF15Xsk1

**Catalogue number:** HSPS00078
**Name:** iTAF15Xsk1
**Description:** Human iPSCs derived from fibroblasts from a patient with Xq24 microdeletion
**Authors:** Menzorov A.G., Nikitina T.V., Tolmacheva E.N., Minaycheva L.I., Nazarenko L.P., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XX, chromosome number variability 46-48, tetraploid cells 3%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** Xq24
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** Xq24 microdeletion, UBE2A deficiency syndrome, recurrent miscarriage, asymmetric X-chromosome inactivation

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 26.08.2022

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF15Xsk4

**Catalogue number:** HSPS00079
**Name:** iTAF15Xsk4
**Description:** Human iPSCs derived from fibroblasts from a patient with Xq24 microdeletion
**Authors:** Menzorov A.G., Nikitina T.V., Tolmacheva E.N., Minaycheva L.I., Nazarenko L.P., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XX, chromosome number variability 46-47, tetraploid cells 5%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** Xq24
**Species control:** cytogenetic
**Cryoconservation passage:** 12
**Area of application:** Xq24 microdeletion, UBE2A deficiency syndrome, recurrent miscarriage, asymmetric X-chromosome inactivation

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 26.08.2022

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:** https://doi.org/10.1134/S1062360423060073

# Cell line passport of iTAF15Xsk6

**Catalogue number:** HSPS00080
**Name:** iTAF15Xsk6
**Authors:** Menzorov A.G., Nikitina T.V., Tolmacheva E.N., Minaycheva L.I., Nazarenko L.P., Lebedev I.N.
**Description:** Human iPSCs derived from fibroblasts from a patient with Xq24 microdeletion
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n=46, XX, chromosome number variability 46-48, tetraploid cells 1%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** Xq24
**Species control:** cytogenetic
**Cryoconservation passage:** 12
**Area of application:** Xq24 microdeletion, UBE2A deficiency syndrome, recurrent miscarriage, asymmetric X-chromosome inactivation

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 26.08.2022

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF15Xsk12

**Catalogue number:** HSPS00082
**Name:** iTAF15Xsk12
**Description:** Human iPSCs derived from fibroblasts from a patient with Xq24 microdeletion
**Authors:** Menzorov A.G., Meshcheryakov N.I., Nikitina T.V., Kashevarova A.A., Tolmacheva E.N., Minaycheva L.I., Nazarenko L.P., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX, chromosome number variability 46-48, tetraploid cells 10%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** Xq24
**Species control:** cytogenetic
**Cryoconservation passage:** 7
**Area of application:** Xq24 microdeletion, UBE2A deficiency syndrome, recurrent miscarriage, asymmetric X-chromosome inactivation

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 30.11.2023

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF15Xsk13

**Catalogue number:** HSPS00083
**Name:** iTAF15Xsk13
**Description:** Human iPSCs derived from fibroblasts from a patient with Xq24 microdeletion
**Authors:** Menzorov A.G., Meshcheryakov N.I., Nikitina T.V., Kashevarova A.A., Tolmacheva E.N., Minaycheva L.I., Nazarenko L.P., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX, chromosome number variability 46-48, tetraploid cells 4%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** Xq24
**Species control:** cytogenetic
**Cryoconservation passage:** 7
**Area of application:** Xq24 microdeletion, UBE2A deficiency syndrome, recurrent miscarriage, asymmetric X-chromosome inactivation

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 30.11.2023

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF15Xsk31

**Catalogue number:** HSPS00084
**Name:** iTAF15Xsk31
**Description:** Human iPSCs derived from fibroblasts from a patient with Xq24 microdeletion
**Authors:** Menzorov A.G., Meshcheryakov N.I., Nikitina T.V., Kashevarova A.A., Tolmacheva E.N., Minaycheva L.I., Nazarenko L.P., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX, chromosome number variability 44-49, tetraploid cells 4%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** Xq24
**Species control:** cytogenetic
**Cryoconservation passage:** 12
**Area of application:** Xq24 microdeletion, UBE2A deficiency syndrome, recurrent miscarriage, asymmetric X-chromosome inactivation

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 30.11.2023

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF15Xsk39

**Catalogue number:** HSPS00085
**Name:** iTAF15Xsk39
**Description:** Human iPSCs derived from fibroblasts from a patient with Xq24 microdeletion
**Authors:** Menzorov A.G., Meshcheryakov N.I., Nikitina T.V., Kashevarova A.A., Tolmacheva E.N., Minaycheva L.I., Nazarenko L.P., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX, chromosome number variability 44-90, tetraploid cells 11%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** Xq24
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** Xq24 microdeletion, UBE2A deficiency syndrome, recurrent miscarriage, asymmetric X-chromosome inactivation

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 30.11.2023

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF5rc11

**Catalogue number:** HSPS00086
**Name:** iTAF5rc11
**Description:** Human iPSCs derived from patient fibroblasts with a ring chromosome 22
**Authors:** Menzorov A.G., Pristyazhnyuk I.E., Nikitina T.V., Kashevarova A.A., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX,r(22) chromosome number variability 46-48, tetraploid cells <1%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** r(22)
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** study of genome instability in the presence of ring chromosomes

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 30.11.2023

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF5rc13

**Catalogue number:** HSPS00087
**Name:** iTAF5rc13
**Description:** Human iPSCs derived from patient fibroblasts with a ring chromosome 22
**Authors:** Menzorov A.G., Pristyazhnyuk I.E., Nikitina T.V., Kashevarova A.A., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX,r(22) chromosome number variability 45-46, tetraploid cells <1%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** r(22)
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** study of genome instability in the presence of ring chromosomes

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 30.11.2023

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF5rc15

**Catalogue number:** HSPS00088
**Name:** iTAF5rc15
**Description:** Human iPSCs derived from patient fibroblasts with a ring chromosome 22
**Authors:** Menzorov A.G., Pristyazhnyuk I.E., Nikitina T.V., Kashevarova A.A., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX,r(22) chromosome number variability 45-46, tetraploid cells 3,3%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** r(22)
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** study of genome instability in the presence of ring chromosomes

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 30.11.2023

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF5rc16

**Catalogue number:** HSPS00089
**Name:** iTAF5rc16
**Description:** Human iPSCs derived from patient fibroblasts with a ring chromosome 22
**Authors:** Menzorov A.G., Pristyazhnyuk I.E., Nikitina T.V., Kashevarova A.A., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX,r(22) chromosome number variability 45-48, tetraploid cells 3,8%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** r(22)
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** study of genome instability in the presence of ring chromosomes

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 30.11.2023

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF5rc17

**Catalogue number:** HSPS00090
**Name:** iTAF5rc17
**Description:** Human iPSCs derived from patient fibroblasts with a ring chromosome 22
**Authors:** Menzorov A.G., Pristyazhnyuk I.E., Nikitina T.V., Kashevarova A.A., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX,r(22) chromosome number variability 45-46, tetraploid cells <1%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** r(22)
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** study of genome instability in the presence of ring chromosomes

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 30.11.2023

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF5rc19

**Catalogue number:** HSPS00091
**Name:** iTAF5rc19
**Description:** Human iPSCs derived from patient fibroblasts with a ring chromosome 22
**Authors:** Menzorov A.G., Pristyazhnyuk I.E., Nikitina T.V., Kashevarova A.A., Lebedev I.N.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46,XX,r(22) chromosome number variability 45-47, tetraploid cells 6,7%, modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** r(22)
**Species control:** cytogenetic
**Cryoconservation passage:** 10
**Area of application:** study of genome instability in the presence of ring chromosomes

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 30.11.2023

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF1-36-H8.1

**Catalogue number:** HSPS00101
**Name:** iTAF1-36-H8.1
**Description:** human iPSCs obtained from fibroblasts of a conditionally healthy donor, introduced deletion of HARsv2_1748 in the *CNTN6* gene (GRCh38/hg38 del3: 1,231,849-1,232,540; 690 bp)
**Authors:** Chvileva A.S., Yunusova A.M., Pristyazhnyuk I.E., Smirnov A.V., Shnaider T.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46, XY, chromosome number variability 46-47, tetraploid cells 8% modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** r(22)
**Species control:** cytogenetic
**Cryoconservation passage:** 12
**Area of application:** studies of HARsv2_1748 deletion, regulatory sequences in the *CNTN6* gene, mental retardation

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 26.08.2022

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:** https://doi.org/10.1134/S1062360424700267

# Cell line passport of iTAF1-36-H8.2

**Catalogue number:** HSPS00102
**Name:** iTAF1-36-H8.1
**Description:** human iPSCs obtained from fibroblasts of a conditionally healthy donor, introduced deletion of HARsv2_1748 in the *CNTN6* gene (GRCh38/hg38 del3: 1,231,849-1,232,540; 690 bp)
**Authors:** Chvileva A.S., Yunusova A.M., Pristyazhnyuk I.E., Smirnov A.V., Shnaider T.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46, XY, chromosome number variability 46-47, tetraploid cells 8% modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:** r(22)
**Species control:** cytogenetic
**Cryoconservation passage:** 12
**Area of application:** studies of HARsv2_1748 deletion, regulatory sequences in the *CNTN6* gene, mental retardation

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 26.08.2022

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 10 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:3 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:** https://doi.org/10.1134/S1062360424700267

# Cell line passport of iTAF1-36-H7.1

**Catalogue number:** HSPS00111
**Name:** iTAF1-36-H7.1
**Description:** human iPSCs obtained from fibroblasts of a conditionally healthy donor, introduced compound heterozygous deletion of HARsv2_1747 in the *CNTN6* gene (GRCh38/hg38 del3: 1,195,873-1,196,314; 442 bp/ del3: 1,195,873-1,196,318; 446 bp)
**Authors:** Knyazeva A.S., Yunusova A.M., Smirnov A.V., Shnaider T.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46, XY, chromosome number variability 46-47, tetraploid cells 5% modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 19
**Area of application:** studies of HARsv2_1747 deletion, regulatory sequences in the *CNTN6* gene, mental retardation

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 26.08.2022

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 20 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:5 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Cell line passport of iTAF1-36-H7.2

**Catalogue number:** HSPS00112
**Name:** iTAF1-36-H7.2
**Description:** human iPSCs obtained from fibroblasts of a conditionally healthy donor, introduced homozygous deletion of HARsv2_1747 in the *CNTN6* gene (GRCh38/hg38 del3: 1,195,873-1,196,314; 442 bp)
**Authors:** Knyazeva A.S., Yunusova A.M., Smirnov A.V., Shnaider T.A.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:** 2n = 46, XY, chromosome number variability 46-47, tetraploid cells 10% modal chromosome number 46
**Pluripotency:** pluripotency demonstrated in embryoid body formation test
**Additional characteristics:**
**Species control:** cytogenetic
**Cryoconservation passage:** 6
**Area of application:** studies of HARsv2_1747 deletion, regulatory sequences in the *CNTN6* gene, mental retardation

**Source**
**Species:** *Homo sapiens*
**Tissue:** fibroblasts
**Date:** 26.08.2022

**Cell culture**
**Morphology:** human ES cell phenotype colonies
**Cell culture method:** monolayer
**Cell culture medium:** DMEM/F12, 20% KSR, NEAA 1%, Glutamine 1%, PenStrep 1%, 2-Mercaptoethanol 0.1 mM, bFGF 20 ng/ml
**Cell culture conditions:** 37°C, 5% $CO_2$
**Passage protocol:** manual passage with the ratio 1:5 - 1:6
**Cryoconservation:** 90% KSR, 10% DMSO
**Cryoconservation cell concentration:** 0.5 mln cells / ml
**Cell viability after cryoconservation:** 5%
**Additional information:** Plastic is coated with 0.1% gelatin. 13.5 dpc fibroblasts of ICR mouse strain are used as feeder cells

**References:**

# Protists

## Cell line passport of THAU1

**Catalogue number:** TAHE00071
**Name:** THAU1
**Description:** the protist *Thraustochytrium aureum* ssp. *strugatskii* was isolated from the dissociated comb jelly *Beroe ovata* (from the Black Sea)
**Authors:** Menzorov A.G., Doroshkov A.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:**
**Pluripotency:**
**Additional characteristics:**
**Species control:** rRNA sequencing
**Cryoconservation passage:** 10
**Area of application:** biotechnology, fatty acid production, study of the Labyrinthulea life cycle

**Source**
**Species:** *Thraustochytrium aureum* ssp. *strugatskii*
**Tissue:**
**Date:** 26.11.2020

**Cell culture**
**Morphology:** “colonies” of cells
**Cell culture method:** monolayer
**Cell culture medium:** a) FAND culture medium: 17 ASW, 5% FBS,
5% DMEM (prepared from powder on 17‰ ASW), x0.05 NEAA, x1 PenStrep; b) 790 By+ (ATCC)
**Cell culture conditions:** room temperature
**Passage protocol:** manual passage (scraping and resuspending) at the ratio 1:10 - 1:100
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 1 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:** http://dx.doi.org/10.7717/peerj.12737

# Cell line passport of THCA1

**Catalogue number:** TAHE00107
**Name:** THCA1
**Description:** the protist *Thraustochytrium caudivorum* was isolated from the biota of the free-living flatworm *Macrostomum lignano*
**Authors:** Menzorov A.G., Biryukov M.Yu.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:**
**Pluripotency:**
**Additional characteristics:**
**Species control:** rRNA sequencing (NCBI Genbank PV862890.1 and PV862891.1)
**Cryoconservation passage:** 12
**Area of application:** study of host-parasite interactions, study of the life cycle of Labyrinthulea

**Source**
**Species:** *Thraustochytrium caudivorum*
**Tissue:**
**Date:** 22.07.2025

**Cell culture**
**Morphology:** single cells
**Cell culture method:** monolayer
**Cell culture medium:** 790 By+ (ATCC)
**Cell culture conditions:** room temperature
**Passage protocol:** manual passage (scraping and resuspending) at the ratio 1:2 - 1:5
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 1 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:**

# Cell line passport of THCA1hygro1

**Catalogue number:** TAHE00108
**Name:** THCA1hygro1
**Description:** the protist *Thraustochytrium caudivorum* was isolated from the biota of the free-living flatworm *Macrostomum lignano*; a DNA fragment pUC19-HygroR-T2A-EGFP containing the hygromycin B resistance gene (*HygroR*) and *EGFP* under the control of the *GAPDH* promoter isolated from *Aurantiochytrium limacinum* was introduced by electroporation
**Authors:** Menzorov A.G., Biryukov M.Yu., Smirnov A.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:**
**Pluripotency:**
**Additional characteristics:** plasmid pUC19-HygroR-T2A-EGFP is based on pUC19_GZG (Addgene, #117226)
**Species control:** rRNA sequencing
**Cryoconservation passage:** 18
**Area of application:** study of host-parasite interactions, study of the life cycle of Labyrinthulea

**Source**
**Species:** *Thraustochytrium caudivorum*
**Tissue:**
**Date:** 22.07.2025

**Cell culture**
**Morphology:** single cells
**Cell culture method:** monolayer
**Cell culture medium:** 790 By+ (ATCC)
**Cell culture conditions:** room temperature
**Passage protocol:** manual passage (scraping and resuspending) at the ratio 1:2 - 1:5
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 1 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:**

# Cell line passport of THCA1zeo3

**Catalogue number:** TAHE00109
**Name:** THCA1zeo3
**Description:** the protist *Thraustochytrium caudivorum* was isolated from the biota of the free-living flatworm *Macrostomum lignano*; a DNA fragment pUC19-GZG-T2A-EGFP containing the blasticidin resistance gene (*shble*) and *EGFP* under the control of the *GAPDH* promoter isolated from *Aurantiochytrium limacinum* was introduced by electroporation
**Authors:** Menzorov A.G., Biryukov M.Yu., Smirnov A.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:**
**Pluripotency:**
**Additional characteristics:** plasmid pUC19-GZG-T2A-EGFP is based on pUC19_GZG (Addgene, #117226)
**Species control:** rRNA sequencing
**Cryoconservation passage:** 18
**Area of application:** study of host-parasite interactions, study of the life cycle of Labyrinthulea

**Source**
**Species:** *Thraustochytrium caudivorum*
**Tissue:**
**Date:** 22.07.2025

**Cell culture**
**Morphology:** single cells
**Cell culture method:** monolayer
**Cell culture medium:** 790 By+ (ATCC)
**Cell culture conditions:** room temperature
**Passage protocol:** manual passage (scraping and resuspending) at the ratio 1:2 - 1:5
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 1 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:**

# Cell line passport of THCA1zeo5

**Catalogue number:** TAHE00110
**Name:** THCA1zeo5
**Description:** the protist *Thraustochytrium caudivorum* was isolated from the biota of the free-living flatworm *Macrostomum lignano*; a DNA fragment pUC19-GZG-T2A-EGFP containing the blasticidin resistance gene (*shble*) and *EGFP* under the control of the *GAPDH* promoter isolated from *Aurantiochytrium limacinum* was introduced by electroporation
**Authors:** Menzorov A.G., Biryukov M.Yu., Smirnov A.V.
**Contamination analysis:** bacteria, fungi and mycoplasma not detected
**Karyotype:**
**Pluripotency:**
**Additional characteristics:** plasmid pUC19-GZG-T2A-EGFP is based on pUC19_GZG (Addgene, #117226)
**Species control:** rRNA sequencing
**Cryoconservation passage:** 18
**Area of application:** study of host-parasite interactions, study of the life cycle of Labyrinthulea

**Source**
**Species:** *Thraustochytrium caudivorum*
**Tissue:**
**Date:** 22.07.2025

**Cell culture**
**Morphology:** single cells
**Cell culture method:** monolayer
**Cell culture medium:** 790 By+ (ATCC)
**Cell culture conditions:** room temperature
**Passage protocol:** manual passage (scraping and resuspending) at the ratio 1:2 - 1:5
**Cryoconservation:** 90% FBS, 10% DMSO
**Cryoconservation cell concentration:** 1 mln cells / ml
**Cell viability after cryoconservation:** 70%
**Additional information:**

**References:**